

UNIVERSITÀ DEGLI STUDI DI MILANO

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[Dario Besusso] CURRICULUM VITAE

INFORMAZIONI PERSONALI (NON INSERIRE INDIRIZZO PRIVATO E TELEFONO FISSO O CELLULARE)

COGNOME	BESUSSO
NOME	DARIO
DATA DI NASCITA	15/01/1976

INSERIRE IL PROPRIO CURRICULUM (non eccedente le 30 pagine)

EDUCATION

- 2002-2007 PhD (Research Doctorate) in Biological Science, Medicine and Surgery Faculty, University of Milan, Italy; Coordinator Prof. Magda Gioia. **Thesis's Title:** Aprataxin and Cell Division (Apratassina e la divisione cellulare).
- 2001 Master Degree in Biology, Faculty of Science at University of Milan with a final evaluation of 110/110.

PROFESSIONAL EXPERIENCES

- 2014-today Senior Postdoc in Elena Cattaneo's lab, Department of Bioscience, University of Milan, Milan, Italy.
- 2013-2014 Research Assistant in Dr. Richard Mellanby's lab, The University of Edinburgh, Queen's Medical Research Institute; Edinburgh, Scotland (UK).
- 2010-2012 Postdoctoral fellow in Liliana Minichiello's laboratory, Centre for Neuroregeneration School of Biomedical Sciences; The University of Edinburgh, Chancellor's Building; Edinburgh, Scotland (UK).
- 2007-2009 Postdoctoral fellow in Liliana Minichiello's laboratory, Mouse Biology Unit, EMBL, Monterotondo (Rome - Italy).
- 2004-2006 Predoctoral fellowship in Jun Qin's laboratory, Department of Biochemistry and Cell Biology at the Baylor College of Medicine, Houston, Texas, (USA).
- 2001-2004 Predoctoral research fellow (AIRC fellowship) for the Molecular Targeting Unit, Department of Experimental Oncology at National Cancer Institute of Milan, (Italy).
- 1999-2001 Studentship in Prof. A. Balsari's laboratory at the National Cancer Institute of Milan, (Italy).

RESEARCH INTERESTS

My research interests for the last decade have been centered on the investigation of pathophysiological mechanisms of neurodegenerative disorders with special focus for Huntington Disease (HD). Since joining Elena Cattaneo's lab, my energies are fully dedicated to HD both in disease mechanisms and development of therapeutic approaches based on human pluripotent stem cells. Moreover, to unleash the full potential of these cell-based systems, I have spent the last few years acquiring and implementing Genome Editing technology for the generation of advanced cell models for HD both for the enhancement of *in vitro* read-outs and cell replacement therapies.

RESEARCH EXPERIENCES

- 2014-today Since joining Prof. Elena Cattaneo's lab at the University of Milan, I have been involved in the NeuroStemCellRepair project aimed to develop cell-based strategies for the treatment of HD. I am also developing approaches of disease modeling and studies of disease mechanisms for HD that are also supported by the American foundation CHDI. I am placing particular effort on applying Genome Editing technologies for the generation of advanced cell models for HD.
- 2012-2013 As a bridge between my past experience in immunity and my present expertise in brain diseases, I have been involved in a project aimed to the *in vitro* and *in vivo* characterization of the role of dendritic cells in the immune response associated with experimentally induced encephalomyelitis (EAE) in the lab of Richard Mellanby at the QMRI, University of Edinburgh, UK.
- 2010-2012 With the relocation of Dr. Liliana Minichiello's lab to the University of Edinburgh, I continued my research about neurotrophin receptors focusing on the role of TrkB in striatal projection neurons. For this reason, I have performed histological, biochemical and behavioral analysis of conditional mouse lines and obtained insights about the role of TrkB in modulating striatopallidal neuron activity. These data are now published in Nat Commun. 2013;4:2031. doi: 10.1038/ncomms3031.
- A second project in the lab was aimed to investigate the role of NGF-TrkA signaling in the postnatal development of the cholinergic system and its relevance to AD. This study is now published in J Neurosci. 2012 Oct 24;32(43):14885-98.
- 2007-2009 Thanks to the acquired experience on protein complex purification, at the beginning of 2007 I was able to join Dr. Liliana Minichiello's laboratory at the Mouse Biology Unit of the EMBL in Rome. Here, I had the opportunity to apply my experience in biochemistry and mass spectrometry to different unique genetic mouse models that allow the *in vivo* identification of protein complexes. In Minichiello's laboratory, that boasts long experience in the field of neurotrophins, I had focused on the purification of the complexes associated with both the neurotrophin receptor TrkB and the protein mutated in Huntington Disease, in different areas of the mouse brain. The aim was to gain insights about the crosstalk of the two proteins in HD thanks to the identification of the different partners by 2D gel electrophoresis and mass spectrometry.
- 2004-2006 After 4 years of immunology cancer research, I moved to United States to acquire specific experience on biochemistry and mass spectrometry. To this aim, I reached the Baylor College of Medicine in Houston where I joined Dr. Jun Qin's laboratory that is specialized on "Network Analysis Proteomics".

Here I acquired extensive experience on biochemistry, protein purification and mass spectrometry technique that I applied into two main projects.

1999-2004 I started my career as student developing anti-cancer therapeutic strategy using bacterial DNA as immune modulator.

Using conventional, knock-out and transgenic mouse models, I have evaluated the contribution of danger sequences to vaccination and to therapy against spontaneous and transplanted tumors. I have also analyzed the mechanism of protection, focusing on the role of the NK-T lymphocyte subset.

I have also analyzed the expression of the receptor TLR9, which respond to bacterial DNA, in mouse intestinal mucosa. I have demonstrated that Paneth cells located in clusters of 5-15 at the bottom of each small intestinal crypt and implicated in antimicrobial activities, express the receptor and degranulate in response to CpG treatment.

COMMUNICATION EXPERIENCES

During my career, I have developed and delivered graduate and undergraduate level course lectures. I have presented original research at departmental and interdepartmental seminars and at national and international congresses. I have participated in researching and writing several grant applications and independently developed fellowship applications. I have also directly and collaboratively developed and written research manuscripts. On daily basis, I review, interpret, and present scientific literature.

PARTICIPATION IN WRITING AND/OR EXECUTING AWARDED GRANT APPLICATIONS

Participation in writing: **Novel Strategies for Cell-based Neural Reconstruction "NSC-Reconstruct" EU funded under Horizon 2020.**

Participation in writing and executing: **ERC 2016 Advanced Grant Proposal (AdG) 742436. HD-DittoGraph: a digital human Embryonic Stem Cell platform for Huntington's repeats.** Funded under H2020-EU.1.1. - EXCELLENT SCIENCE - European Research Council (ERC)

Participation in writing and executing: **PRIN 2015. Generation of functional striatal neurons for brain repair in Huntington Disease.** Funded under the national network managed by the Ministry of Education, University and Research of the Italian government.

Participation in writing and executing: **CHDI Foundation, Joint Steering Committee 2016-2019.** CHDI Foundation is a privately-funded, not-for-profit biomedical research organization devoted to Huntington's disease. The mission is to speed-up the development of effective therapeutics for HD.

Participation in executing: **NeuroStemcellRepair.** The European stem cell consortium for neural cell replacement, reprogramming and functional brain repair is a 4-year FP7 European Union funded Project which has been awarded a total budget of 6 million EURO and is led by Coordinator Prof. Elena Cattaneo – University of Milan and Deputy Coordinator Prof. Ernest Arenas – Karoliska Institutet.

Participation in executing: **European Sixth Framework Program** 2002-2006, EU FP6
MEMORIES, 037831

ACADEMIC ACTIVITIES

Attività didattiche integrative e compiti didattici extra-curricolari. 562/AB-tutorato per: Tirocinio percorso 11; Neurobiologia-Modulo 1; Responsabile: Prof.ssa Chiara Zuccato. 11-19 March 2019. University of Milano, Department of Bioscience, Milano, Italy.

Didactic lesson: “Genome Editing: history, mechanisms and applications.” 16 Nov 2018, Tecniche Avanzate di Indagine Biomedica, University of Milano, Department of Bioscience, Milano, Italy.

Attività didattiche integrative e compiti didattici extra-curricolari. 373/AB-tutorato per: Tirocinio percorso 11; Neurobiologia-Modulo 1; Responsabile: Prof.ssa Chiara Zuccato. 26 Feb-9 March 2018. University of Milano, Department of Bioscience, Milano, Italy.

Didactic lesson: “Genome editing and applications to hPSCs”. 16 May 2017, University of Milano, Department of Bioscience, Milano, Italy.

UNISTEM DAY 2015. Dr. Dario Besusso, Centro di Ricerca sulle Cellule Staminali – Università degli Studi di Milano. “Riparare il cervello: cellule staminali e non solo”. Varese, 13 March, 2015.

Neuroscience Seminar Series 2012. Centre for Neuroregeneration (CNR) – Edinburgh University, Biomedical Science “Huntington Disease and BDNF signaling”, 23 May, 2012.

COURSE

Hydra X - The European Summer School on Stem Cell & Regenerative Medicine. Hydra, Greece 7 - 14, Settembre 2014.

SUPERVISION OF STUDENT’S DISSERTATIONS

Sara Oppi

Thesis title: *Differenziamento a neuroni striatali di cellule staminali embrionali “Clinical Grade”*. 2016. Final grade of 110/110 cum laude.

Manuel Cernigoj

Thesis title: *Strategie per la Riprogrammazione di fibroblasti a neuroni striatali come modello di malattia di Huntington*. 2017. Final grade of 110/110 cum laude.

Andrea Cossu

Thesis title: *Generazione di linee hES per l’arricchimento di progenitori striatali in vitro per trapianto sperimentale nel modello Huntington*. 2019. Final grade of 110/110 cum laude.

- **Genome Editing**
 - Excellent knowledge on the use of CRISPR/Cas9/Cpf1 editing strategies to achieve target-specific KO or KI of exogenous DNA fragments in active or silent coding sequences both in human and mouse cell models.
- **Immunohistochemistry**
 - Extensive experience on fluorescent and colorimetric immunohistochemistry in neuronal tissue or *in vitro* cultures.
- **Molecular Biology**
 - Cloning of constructs for liposome-mediated transient/stable expression of fusion proteins in primary or stable cell lines;
 - Cloning of constructs for virus-mediated stable expression of fusion proteins in primary or stable cell lines;
 - Gene silencing using iRNA and virus-transduced shRNA in primary or stable cell lines;
 - PCR; RT-PCR; Quantitative Real-Time PCR with SYBR Green or probe-based system;
 - Southern e Northern blotting.
 - Nanopore DNA sequencing.
- **Cellular Biology**
 - Cell cultures in sterile condition with immortalized and primary cell lines derived from different area of the brain, spleen, bone marrow.
 - Long term maintenance of primary and stable cell lines
 - Proliferative and cytotoxic assays; colony assay.
- **Biochemistry**
 - SDS-PAGE and related techniques; WB, IP, Co-IP and Large Scale IP for Mass-Spec Analysis.
 - Two-Dimensional (2D) gel electrophoresis for comparative proteomic analysis.
- **Immunology**
 - ELISA, ELISpot.
 - Cytometry: characterisation of the expression of cellular markers; assays to analyze apoptosis, cell cycle and related check-point, proliferation, cytokine production with multiparametric analysis using BD FACScalibur (CellQuest, PaintaGate, WinMDI) and Backman-Coulter ELISE instrument.
- **Microscopy**
 - Multicolor conventional laser scanning confocal microscopy (tissue and cell culture) and applied to fluorescence recovery after photobleaching (FRAP) and fluorescence resonance energy transfer (FRET) analysis;
 - Conventional light microscopy for colocalisation studies by fusion proteins or conventional immunofluorescence.
 - Time laps live cell imaging. Digital imaging and all the analysis related to image processing and video editing.
- **Experience with animals**
 - Extensive experience on mouse handling, colony management and *in vivo* procedures (injections of substances by i.p., i.v. or s.c.; retrorbital bleeding, tissue dissection, tagging, etc.)
 - Whole mouse fixation via transcordial perfusion
- **Animal Behaviour**

- Extensive experience on different behavioural tasks using mouse models in order to address memory/learning, motor coordination/balance, fear condition, drug addiction/relapse, and exploration.
- **Computer Skills**
 - Excellent use of Microsoft Office package including Word, Excel, Access, Powerpoint. Knowledge of basic element of Unix-Linux. Use of DNA/RNA and protein analysis software. Imaging and movie processing (ImageJ, Photoshop, Illustrator).
 - Good knowledge of statistical applications like StatView, Excel and Prism.
 - Software for the 3D reconstruction of neuronal cells and relative analysis (Bitplane, Imaris). Experience with NeuroLucida and other proprietary software like ZEN, AxioVision, and Velocity.

PUBLICATIONS

1. **Besusso D.**, Cernigoj M., Cossu A., Conforti P., Trovesi C., Cattaneo E. A CRISPR strategy to generate multifunctional reporters for the subpallial determinant GSX2. **Submitted.**
2. Wang Y., Bel L., Russell C., Armstrong C., Wu Y., Spampanato J., Cui J., Tarboton P., **Besusso D.**, Vezzoli E., Das S., Coon H., Shecheglovitov, A. Human cortical organoids from single iPSC-derived neural rosettes reveal a novel subtype of FOXP2 inhibitory neurons and early functional deficits induced by SHANK3 deficiency. **Submitted.**
3. **Besusso D.**, Schellino R., Boido M., Belloli S., Parolisi R., Conforti P., Faedo A., Cernigoj M., Campus I., Laporta A., Dickinson Bocchi V., Murtagh V., Parmar M., Spaiardi P., Talpo F., Maniezzi C., Toselli M.G., Biella G., Moresco R.M., Vercelli A., Buffo A. and Cattaneo E. Stem cell derived human striatal progenitors innervate striatal targets and alleviate sensorimotor deficit in a rat model of Huntington Disease. **Stem Cell Reports, 2020 May 12;14(5):876-891. doi: 10.1016/j.stemcr.2020.03.018. IF:5.499.**
4. Vezzoli E., Caron I., Talpo F., **Besusso D.**, Conforti P., Battaglia E., Sogne E., Falqui A., Petricca L., Verani M., Martufi P., Caricasole A., Bresciani A., Cecchetti O., Rivetti di Val Cervo P., Sancini G., Riess O., Phuc Nguyen H., Seipold L., Saftig P., Biella G., Cattaneo E., Zuccato C.. Inhibiting pathologically active ADAM10 rescues synaptic and cognitive decline in Huntington's disease. **J Clin Invest. 2019 May 6;130. doi: 10.1172/JCI120616. IF:13.25**
5. Mair I, **Besusso D**, Saul L, Patel SD, Ravindran R, McPherson RC, Leech MD, O'Connor RA, Anderton SM, Mellanby RJ. PD-1 expression is upregulated on adapted T cells in experimental autoimmune encephalomyelitis but is not required to maintain a hyporesponsive state. **Eur J Immunol. 2019 Jan;49(1):112-120. doi: 10.1002/eji.201847868. IF 4.247.**
6. Conforti P, **Besusso D**, Bocchi VD, Faedo A, Cesana E, Rossetti G, Ranzani V, Svendsen CN, Thompson LM, Toselli M, Biella G, Pagani M, Cattaneo E. Faulty neuronal determination and cell polarization are reverted by modulating HD early phenotypes. **Proc Natl Acad Sci U S A. 2018an 23;115(4):E762-E771. doi: 10.1073/pnas.1715865115. IF: 9.661.**
7. Faedo A, Laporta A, Segnali A, Galimberti M, **Besusso D**, Cesana E, Belloli S, Moresco RM, Tropiano M, Fucà E, Wild S, Bosio A, Vercelli AE, Biella G, Cattaneo E.

- Differentiation of human telencephalic progenitor cells into MSNs by inducible expression of Gsx2 and Ebf1. *Proc Natl Acad Sci U S A*. 2017 Jan 30. pii: 201611473. doi: 10.1073/pnas.1611473114. IF: 9.423.
8. Saul L, **Besusso D**, Mellanby RJ. LPS-matured CD11c+ bone marrow-derived dendritic cells can initiate autoimmune pathology with minimal injection site inflammation. *Lab Anim*. 2017 Jun;51(3):292-300. doi: 10.1177/0023677216663584. IF: 1.450
 9. **Besusso D.**, Saul L., Leech M.D., O'Connor R.A., MacDonald A.S., Anderton S.M., Mellanby R.J. 1,25-Dihydroxyvitamin D3-Conditioned CD11c+ Dendritic Cells are Effective Initiators of CNS Autoimmune Disease. *Front Immunol*. 2015 Nov 18;6:575. doi: 10.3389/fimmu.2015.00575. IF:5.695.
 10. **Besusso D.**, Geibel M., Kramer D., Pendolino V., Picconi B., Calabresi P., Minichiello L. Depletion of BDNF/TrkB signaling in striatopallidal neurons alters inhibition of locomotor behaviour. *Nat Commun*. 2013;4:2031. doi: 10.1038/ncomms3031. IF:11.329.
 11. Müller M.*, Triaca V.*, **Besusso D.***, Costanzi M., Horn JM., Koudelka J., Geibel M., Cestari V., and Minichiello L. Loss of NGF-TrkA Signaling From The Central Nervous System Is Not Sufficient To Induce Cognitive Impairments in Young-Adult or Intermediate-Aged Mice. *J Neurosci*. 2012 Oct 24;32(43):14885-98, doi: 10.1523/JNEUROSCI.2849-12.2012. * M.M., V.T. and D.B. contributed equally to this work. IF: 5.924.
 12. Leng M, **Besusso D**, Jung SY, Wang Y, Qin J. Targeting Plk1 to chromosome arms and regulating chromosome compaction by the PICH ATPase. *Cell Cycle*. 2008 May 15;7(10):1480-9, doi: 10.4161/cc.7.10.5951. IF: 3.952.
 13. Mu JJ, Wang Y, Luo H, Leng M, Zhang J, Yang T, **Besusso D**, Jung SY, Qin J. A proteomic analysis of ataxia telangiectasia-mutated (ATM)/ATM-Rad3-related (ATR) substrates identifies the ubiquitin-proteasome system as a regulator for DNA damage checkpoints. *J Biol Chem*. 2007 Jun 15;282(24):17330-4, doi: 10.1074/jbc.C700079200. IF: 4.573.
 14. Sfondrini L, Rossini A, **Besusso D**, Merlo A, Tagliabue E, Menard S, Balsari A. Antitumor Activity of the TLR-5 Ligand Flagellin in Mouse Models of Cancer. *J Immunol*. 2006 Jun 1;176(11):6624-30, doi: 10.4049/jimmunol.176.11.6624. IF: 5.95
 15. Rumio C*, **Besusso D***, Arnaboldi F, Palazzo M, Selleri S, Gariboldi S, Akira S, Uematsu S, Bignami P, Ceriani V, Menard S, Balsari A. Activation of smooth muscle and myenteric plexus cells of jejunum via toll-like receptor 4. *J Cell Physiol*. 2006 Jul;208(1):47-54, doi: 10.1002/jcp.20632. * C.R. and D.B. contributed equally to this work. IF:3.839.
 16. Rumio C.*, **Besusso D.***, Palazzo M., Selleri S., Ménard S. and Balsari A.: "Paneth cells express Toll-like Receptor 9 and degranulate after exposure to bacterial DNA". *Am. J. Pathol.*, 2004 Aug;165(2):373-81, doi: 10.1002/jcp.20360, doi: 10.1016/S0002-9440(10)63304-4. * C.R. and D.B. contributed equally to this work. IF:6.967.
 17. Nardini E, Morelli D, Aiello P, **Besusso D**, Calcaterra C, Mariani L, Palazzo M, Vecchi A, Paltrinieri S, Menard S, Balsari A. CpG-oligodeoxynucleotides induce mobilization of hematopoietic progenitor cells into peripheral blood in association with mouse KC (IL-8) production. *J Cell Physiol*. 2005 Sep;204(3):889-95, doi: 10.1002/jcp.20360. IF: 3.839.
 18. Battaini F, **Besusso D**, Sfondrini L, Rossini A, Morelli D, Tagliabue E, Menard S, Balsari A. Antibody response after vaccination with antigen-pulsed dendritic cells. *Int J Biol Markers*. 2004 Jul-Sep;19(3):213-20. IF: 0.92

19. Balsari A., Tortoreto M., **Besusso D.**, Petrangolini G., Sfondrini L., Maggi R., Ménard S. and Pratesi G. “Combination of a CpG-oligodeoxynucleotide and a topoisomerase I inhibitor in therapy of human tumor xenografts.” **Eur. J. Cancer**, 2004 May;**40(8):1275-81**, doi: **10.1016/j.ejca.2004.01.023**. IF: 5.417.
20. Sfondrini L., **Besusso D.**, Bronte V., Macino B., Colombo M.P., Ménard S., Balsari A.: “CpG-Oligodeoxynucleotides activate tyrosinase-related protein 2-specific T lymphocytes but do not lead to a protective tumor-specific memory response”. **Cancer Immunol. Immunother.**, 2004 Mar; **18**, doi: **10.1007/s00262-004-0516-x**. IF: 4.086.
21. Sfondrini L., **Besusso D.**, Rumio C., Rodolfo M., Ménard S. and Balsari A.: “Prevention of spontaneous mammary adenocarcinoma in Her-2/neu transgenic mice by CpG-oligodeoxynucleotides”. **FASEB Journal**. 2002 Nov; **16(13):1749-54**, doi: **10.1096/fj.02-0383com**. IF: 5.299.
22. Sfondrini L., **Besusso D.**, Zoia M.T., Rodolfo M., Invernizzi A.M., Taniguchi M., Nakayama T., Colombo M.P., Ménard S. and Balsari A.: “Absence of the CD1 molecule upregulates anti-tumor activity induced by CpG oligodeoxynucleotides in mice”. **J Immunol**. 2002 Jul 1; **169(1):151-8**, doi: **10.4049/jimmunol.169.1.151**. IF: 4.920.

Total average IF: 5.6831

Top10 average IF: 7.6868

ORAL PRESENTATIONS

1. **Gene- and Cell-Based Therapies: CRISPR, Stem Cells, and Beyond** “ESC-based cell therapy in a preclinical model of Huntington disease” 2-4 March 2020; San Francisco, CA, USA.
2. **17th SINS National Congress**. Harnessing CRISPR/Cas9 technology to generated stem cell-based model of Huntington disease. 01-04 October 2017 Ischia, Italy.
3. **Neurostemcellrepair Workshop**, Progress in hES-based cell therapy for HD. 6-8 Novembre 2016, Roma, Italy.
4. **III° Neurostemcellrepair Annual Meeting**, Progress in hES-based cell therapy for HD. 9-11 April 2016, Bellagio. Italy.
5. **Neurostemcellrepair Workshop & Public Event** at the Italian Republic Senate, Progress in hES-based cell therapy for HD. 8-10 Novembre 2015, Roma.
6. **II° Neurostemcellrepair Annual Meeting**, Progress in hES-based cell therapy for HD. 11-13 April 2015, Bellagio. Italy.

POSTER ABSTRACTS

1. Conforti P., **Besusso D.**, Campus I., Bocchi V., Cattaneo E. An optimized protocol for efficient and reproducible production of hESC-derived striatal projection neurons. 13th Annual Huntington's Disease Therapeutics Conference: A Forum for Drug Discovery & Development. Feb 26-Mar 1, 2018. The Parker Palm Springs Hotel, Palm Springs, California, USA.

2. **Besusso D.**, Cernigoj M., Laporta A., Cattaneo E. Multicolor CRISPR labeling of key striatal fate determinants in human embryonic stem cells. **CHDI's 12th Annual HD Therapeutics Conference** April 24 – 27, 2017, Malta.
3. **Besusso D.**, Cernigoj M., Laporta A., Cattaneo E. Multicolor CRISPR labeling of key striatal fate determinants in human embryonic stem cells. **Genome Editing in Neuroscience**, 22nd April 2016, Paris. France.
4. **Besusso D.**, Belloli S., Di Grigoli G., Moresco R.M., Tropicano M., Vercelli A., Cattaneo E. Transplantation of human pluripotent stem cells-derived MSN progenitors in animal models of Huntington disease. **II° Neurostemcellrepair Annual Meeting**, 11-13 April 2015, Bellagio. Italy.
5. **Besusso D.**, Lancioni A, Motta B, Cattaneo E. Direct conversion of mouse fibroblasts into striatal MSNs. **Hydra X - The European Summer School on Stem Cell & Regenerative Medicine**. 7-14 September 2014, Hydra, Greece.
6. **Besusso D.**, Geibel M., Kramer D., Pendolino V., Picconi B., Calabresi P., Minichiello L. **“BDNF-TrkB Signaling in Striatopallidal Neurons Controls Inhibition of Locomotor Behaviour”**. Neuroscience Day 2012, March 21th 2012; Edinburgh, UK.
7. **Besusso D.** and Minichiello L. **“BDNF/TrkB signaling is not required for Medium-sized Spiny Neurons survival and maintenance”**. NGF 2010, Neurotrophic factors in health and disease; June 10-13, 2010; Helsinki, Finland.
8. **Besusso D.** and Minichiello L. **“Conditional deletion of TrkB from enkephalin-expressing neurons does not lead to Huntington-like neuronal degeneration”**. Cortical Interneurons in Health and Disease; 21 - 25 June | 2009 |Costa d'en Blanes | Mallorca| Spain.
9. Sfondrini L., **Besusso D.**, Rodolfo M., Ménard S. and Balsari A. **“Therapeutic effect and tumor prevention by CpG oligonucleotides”** 1st National Conference SIICA, Montecatini Terme, May 8-11, 2002. Conference Abstract in *Minerva Biotechnologica* 14 (1): 60 (2002).
10. Sfondrini L., **Besusso D.**, Bronte V., Macino B., Colombo M.P., Ménard S., Balsari A.. **”Induction of a lymphocyte response specific to Tyrosinase-Related Protein 2 by CpG treatment of B16 melanoma bearing mice”** 15th European Immunology Congress (EFIS 2003), Jun 8-12 2003, Rhodes (Greece), poster session.
11. Sfondrini L., **Besusso D.**, Bronte V., Macino B., Colombo M.P., Ménard S., Balsari A.. **“CpG-oligodeoxynucleotides activate Tyrosinase-Related Protein 2 specific T lymphocytes but does not lead to a protective tumor-specific memory response”** XLV National Congress of Italian Cancer Society (SIC), Nov. 9-12, 2003; Bergamo (Italy), poster session.

CONGRESS PARTICIPATIONS

1. **13th Annual Huntington's Disease Therapeutics Conference: A Forum for Drug Discovery & Development**. Feb 26-Mar 1, 2018. The Parker Palm Springs Hotel, Palm Springs, California, USA.
2. **17th SINS National Congress**. Harnessing CRISPR/Cas9 technology to generated stem cell-based model of Huntington disease. 01-04 October 2017 Ischia, Italy.
3. **CHDI's 12th Annual HD Therapeutics Conference** April 24 – 27, 2017, Malta.
4. **Neurostemcellrepair Workshop**, 6-8 Novembre 2016, Roma. Italy.
5. **Genome Editing in Neuroscience**, 22nd April 2016, Paris. France.
6. **III° Neurostemcellrepair Annual Meeting**, 9-11 April 2016, Bellagio. Italy.

7. **VI Meeting on the Molecular Mechanisms of Neurodegeneration**, 28-30, Maggio 2015, Milano.
8. **Multi-site Differentiation Consortium 45 month JSC Meeting**, 19-20 ottobre 2015, Milano
9. **Neurostemcellrepair Workshop & Public Event** at the Italian Republic Senate, 8-10 Novembre 2015, Roma.
10. **Multi-site Differentiation Consortium 39 month JSC Meeting**, 16-17 Aprile 2015, Barcellona.
11. **II° Neurostemcellrepair Annual Meeting**, 11-13 April 2015, Bellagio. Italy.
12. **Hydra X - The European Summer School on Stem Cell & Regenerative Medicine**. 7-14 September 2014, Hydra, Greece.
13. **I° Neurostemcellrepair Annual Meeting**, 10-12 April 2014, Bellagio. Italy.
14. **Neuroscience Day 2012**, March 21th 2012; Edinburgh, UK.
15. **NGF 2010, Neurotrophic factors in health and disease**; June 10-13, 2010; Helsinki, Finland.
16. **Cortical Interneurons in Health and Disease**; 21 - 25 June | 2009 |Costa d'en Blanes | Mallorca| Spain.
17. **1st National Conference SIICA**, Montecatini Terme, May 8-11, 2002. Conference Abstract in *Minerva Biotecnologica* 14 (1): 60 (2002).
18. **XLV National Congress of Italian Cancer Society (SIC)**, Nov. 9-12, 2003; Bergamo (Italy), poster session.

REFERENCES

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Reference letters can be obtained upon request.

I hereby grant permission to use my personal data in accordance with [Art. 13, Legislative Decree n. 196/03 - "Personal Data Protection Code"](#)

Data

07/07/2020

Luogo

Milano